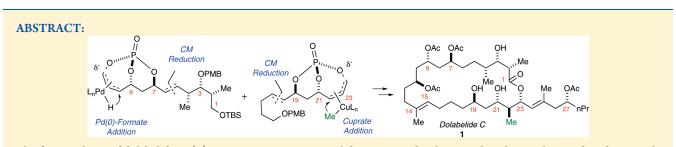
Total Synthesis of Dolabelide C: A Phosphate-Mediated Approach

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S Supporting Information



The first synthesis of dolabelide C (1), a cytotoxic marine macrolide, is reported utilizing a phosphate tether-mediated approach. Bicyclic phosphates (S,S,S_P) -5 and (R,R,R_P) -5 serve as the central building blocks for the construction of two major 1,3-anti-diol subunits in 1 through selective cleavage pathways, regioselective olefin reduction, and cross-metathesis. Overall, phosphatemediated processes provided copious amounts of both major subunits allowing for a detailed RCM macrocyclization study to the 24-membered macrolactone 1.

INTRODUCTION: THE DOLABELIDE FAMILY

In 1995, the isolation and structural characterization of two new 22-membered macrolides, dolabelides A and B, from the sea hare Dolabella auricularia was reported (Figure 1). These compounds exhibited cytotoxicity against cervical cancer HeLa-S3 cells with IC50 values of 6.3 and 1.3 μ g/mL, respectively.¹ Two years later, dolabelides C and D,² 24-membered macrolides, were isolated from the same source and were found to possess cytotoxicity toward HeLa-S₃ cells with IC₅₀ values of 1.9 and 1.5 μ g/mL, respectively. To the best of our knowledge, the mechanism of action of these compounds remains unknown to date.

Common features among the dolabelide family are 11 stereogenic centers, eight of which bear oxygen, and two E-configured trisubstituted olefins. Other structural features possessed by this family of macrolactones include 1,3-anti-diol fragments found at C7/C9 and C19/C21, along with an accompanying 1,3-syn-diol at C9/C11 and polypropionate fragments at C1/C4 and C21/ C23. The stereochemical complexity and biological profile of this class of compounds has attracted synthetic interest from several groups,³ and in 2006, the first total synthesis of dolabelide D was reported by Leighton and co-workers.⁴

Dolabelide C (1) can be disconnected into C1-C14 and C15-C30 subunits, 2 and 3, respectively (Scheme 1). The endgame for this approach is similar to Leighton's strategy toward dolabelide D,⁴ employing a macrocyclization sequence to install the C14/C15 trisubstituted olefin through a late-stage ring-closing metathesis (RCM) reaction. Macrocyclization, via RCM, is preceded by Yamaguchi coupling between the C1 carboxylic acid of the northern subunit 2 and the C23 carbinol center in the southern subunit 3. Central to this approach are the 1,3 anti-diol motifs at C7/C9 and C19/C21, which can be

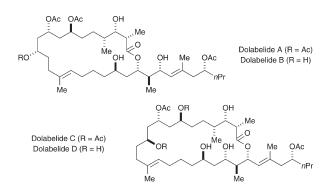


Figure 1. Dolabelide family, isolated from Dollabella auricularia.

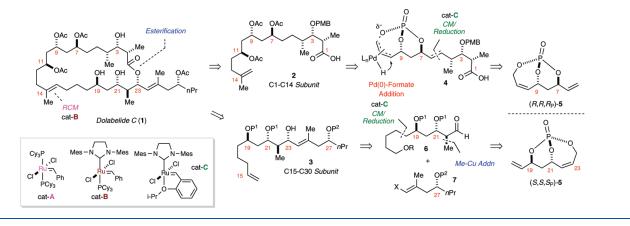
assembled and elaborated from bicyclic phosphates (R,R,R_P) -5 and (S_1, S_2, S_P) -5, respectively, utilizing a phosphate tethermediated approach.

RESULTS AND DISCUSSION

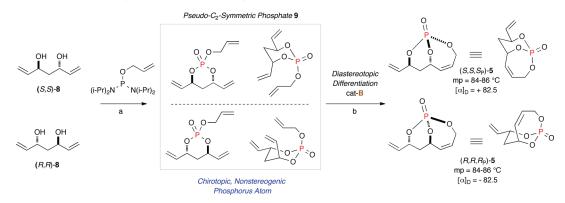
I. Construction of P-Chiral, Nonracemic Bicyclo[4.3.1]phosphates (*R*,*R*,*R*_P)-5 and (*S*,*S*,*S*_P)-5. The enantiomeric phosphate triester building blocks (R_1, R_2, R_P) -5 and (S_1, S_2, S_P) -5 (Scheme 2) were assembled via a phosphate tether, RCM desymmetrization approach⁵ inspired by Burke and co-workers.⁶ In this method, a phosphate tether effectively serves to mediate the tripodal coupling of *anti*-diol 8^7 with an allylic alcohol component via either a phosphoryl monochloride or through a

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Scheme 1. Retrosynthesis of Dolabelide C



Scheme 2. Tether-Mediated Desymmetrization of C2-Symmetric 1,3-anti-Diol 8^a



^{*a*} Reagents and conditions: (a) allyl tetraisopropylphosphorodiamidite, 1*H*-tetrazole, MeCN, 2 h, rt, then *m*-CBPA, 1 h, 64%; (b) cat-**B**, CH₂Cl₂, 85–90%.

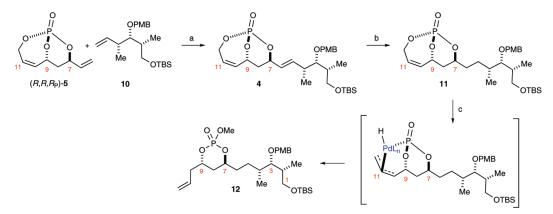
one-step coupling/oxidation sequence from commercially available allyl tetraisopropylphosphorodiamidite to yield pseudo- C_2 -symmetric triene **9**. Desymmetrization⁸ by ring-closing metathesis (RCM) using Grubbs catalyst [(IMesH₂)(PCy₃)(Cl)₂Ru=CHPh (cat-B)]⁹⁻¹¹ affords *P*-chiral bicyclo[4.3.1]phosphate (*S*,*S*,*S*_P)-**5** and is based on the premise that only the terminal olefin *cis* to the phosphate-tethered olefin reacts to generate **5** possessing two sterically differentiated olefins.

II. Construction of C1–C14 Subunit. We embarked upon the synthesis of the C1–C14 subunit of dolabelide beginning with an elaborate cross-methesis (CM) between bicyclic phosphate (R,R,R_p) -5 possessing a type III exocyclic terminal olefin and type I olefin **10**.¹² As shown previously,¹³ CM of (R,R,R_p) -5 is high yielding with both type I and type II olefins in the presence of the Hoveyda–Grubbs catalyst (cat-C).¹⁴ Various conditions for the desired CM of **10** and (R,R,R_p) -5 were probed, and it was found that employing 6 mol % of Hoveyda–Grubbs catalyst at 90 °C in DCE gave the CM adduct 4 in 72% yield (Scheme 3). This notable CM event between two complex olefins assembles in a single operation five of the six stereocenters contained within the C1–C14 subunit of dolabelide C. Moreover, excess amounts of **10**, a type II CM partner, could be recovered in near-quantitative yield and recycled in future CM events.

Two related regioselective processes were next investigated. The first involved a regioselective removal of the exocyclic C5–C6 olefin in the CM adduct 4 in the presence of the C10–C11 internal olefin, which sets the stage for subsequent regioselective hydride opening of the bicyclic system. Upon probing several hydrogenation conditions (Wilkinson's catalyst, Crabtree's catalyst, Pd/C), it was found that a mild diimide reduction, generated in situ from *o*-nitrobenzenesulfonyl hydrazine,¹⁵ provided the necessary hydrogenated phosphate moiety **11** with near-complete regioselectivity for the exocyclic olefin. In comparison, other diimide conditions (tosylhydrazine, NaOAc, H₂O, DCE, 90 °C) gave drastically lower yields, likely due to bicyclic phosphate instability under basic medium.¹⁶

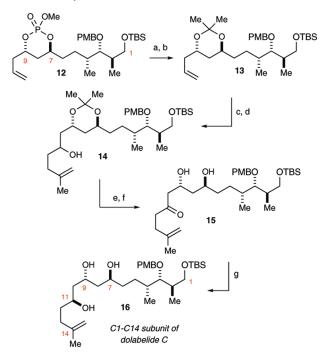
Having achieved the regioselectively hydrogenated product **11**, a regioselective opening with hydride was probed as an additional method to unmask the phosphate tether. Initial studies focused on allylic copper hydride addition using various reagents (Stryker's reagent, CuCN \cdot 2LiCl/PhSiH₃, CeCl₃ \cdot 7H₂O/NaBH₄) (Scheme 3). Unfortunately, all conditions probed provided only unreacted starting material or total decomposition of the reaction mixture. Pd-catalyzed formate reductions were next investigated for generation of the requisite terminal olefin.¹⁷ Employment of 1.5 equiv of formic acid and 5 mol % of Pd(OAc)₂ at 40 °C in DCE selectively opened phosphate **11** to provide the desired terminal olefin in **12**. Methylation of the phosphate acid intermediate showed that a highly

Scheme 3. Phosphate-Mediated Sequence for Assembly of the C1–C14 Subunit^a



^{*a*} Reagents and conditions: (a) cat-C (6 mol %), DCE, 90 °C, 72%; (b) o-NO₂C₆H₅SO₂NHNH₂, Et₃N, CH₂Cl₂, 72%; (c) Pd(OAc)₂ (5 mol %), HCO₂H, Et₃N, DCE, 40 °C, then MeOH, TMSCHN₂, 87%. Abbreviations: DCE = 1,2-dichloroethane.

Scheme 4. Synthesis of C1–C14 Carbon Framework^a



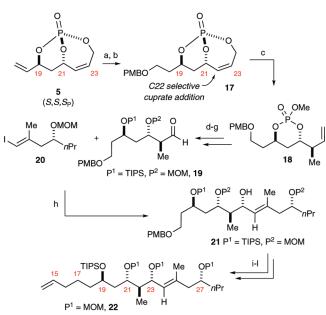
^a Reagents and conditions: (a) LiAlH₄, Et₂O, 75%; (b) PPTS, 2,2-DMP, CH₂Cl₂, 96%; (c) O₃, pyridine, 1:1 MeOH/CH₂Cl₂, -78 °C, then Me₂S, 72%; (d) 1-iodo-3-methylbutene, Mg, Et₂O, -78 °C, 95%; (e) Dess-Martin periodinane, NaHCO₃, CH₂Cl₂, 90%; (f) CeCl₃·7H₂O, H₂O/MeCN (1:7), 87%; (g) Et₂BOMe, NaBH₄, THF/MeOH 4:1, -78 °C, ds \geq 20:1, 60% (95% brsm).

regioselective process was operative (>20:1 ratio of regioisomers as evident by ³¹P NMR analysis). Purification provided phosphate **12** in 87% yield. The remarkable regioselectivity reveals another feature of the phosphate tether, whereby orthogonal orbital alignment within **11** allows for selective Pd(0)-catalyzed ionization of C12 over the C9 allylic phosphate position. This ionization allows Pd to deliver the hydride selectively at the internal C10 position to provide the desired terminal olefin. Addition of the hydride at the terminal C12 position would afford an allylic phosphate anion that is capable of additional ionization events with the C9 phosphate. Ultimately, the success of this reaction results in a net olefin transposition expediting the route to the C1-C14 subunit, as well as showcasing an additional facet of the phosphate-mediated methodology.

Upon completion of the synthesis of phosphate 12, work began toward the installation of the C11-C14 fragment (Scheme 4). Cleavage of the phosphate was achieved using LiAlH₄, which generated a diol that was subsequently protected (PPTS, 2, 2-dimethoxypropane, CH₂Cl₂) to yield acetonide 13. Subsequent ozonolysis (O₃, pyridine, CH₂Cl₂/MeOH 1:1, Me₂S) of the terminal olefin produced the requisite aldehyde, which was subjected to the corresponding Grignard¹⁸ derived from 1-iodo-3-methyl-3-butene affording 14 in a 95% yield. Dess-Martin periodinane (DMP, NaHCO₃, CH₂Cl₂) oxidation of the free alcohol in 14 produced the corresponding ketone in 90% yield. Attempts to selectively reduce the acetonide-protected ketone, using an assortment of reducing agents, failed to give any diastereoselectivity at C11. This problem was circumvented by deprotection of the acetonide and subsequent syn reduction utilizing the C9 alcohol. Selective removal of the acetonide was achieved by the addition of CeCl₃·7H₂O and water,¹⁹ which efficiently (86% yield) cleaved the acetonide-protecting group without loss of the primary TBS group to provide diol 15. Final reduction of ketone 15 using Evan's conditions $(Et_2BOMe, NaBH_4)^{20}$ afforded triol 16 in 60% (95% brsm) with excellent diastereoselectivity (ds \geq 20:1).

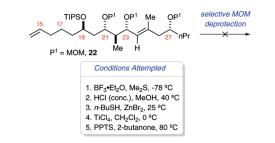
III. First-Generation Synthesis of the C15–C30 Subunit of Dolabelide C. The construction of the C15–C30 subunit of dolabelide began with the enantiomeric bicyclic phosphate (S,S,S_P) -5 possessing unique orbital symmetry of the bicyclic phosphate (Scheme 5).²¹ Initial studies probed the possibility of an oxidation of the exocyclic olefin of **5** in the presence of the cyclic olefin. After testing various conditions, a chemoselective hydroboration of (S,S,S_P) -5 was achieved using 9-BBN followed by mild NaBO₃ oxidation to yield the primary alcohol. Because of the aforementioned instability of (S,S,S_P) -5 to basic hydrolysis, a mild perborate oxidation protocol developed by Burke and co-workers was implemented. Burke has shown this protocol to be compatible with multiple acetate protecting groups;²²

Scheme 5. First-Generation Synthesis of the C15–C30 Subunit^a



^a Reagents and conditions: (a) 9-BBN, then H_2O , NaBO₃·4 H_2O , 80%; (b) *p*-OMeC₆ $H_4OCH_2OC=$ NH(CCl₃), PPTS, CH₂Cl₂, 89%; (c) (1) CuCN·2LiCl, Me₂Zn, THF, -30 °C to rt; (2) TMSCHN₂, MeOH, 87%; (d) LiAlH₄, Et₂O, 0 °C, 96%; (e) TIPSCl, imidazole, DMAP, CH₂Cl₂, 86%; (f) MOMCl, iPr₂NEt, CH₂Cl₂, 91%; (g) O₃, pyridine, -78 °C, Me₂S, 75%; (h) *t*-BuLi, ZnBr₂, **20**, then *n*-BuLi, (*R*,*S*,)-NME, then **19**, 65%, 11:1 dr; (i) MOMCl, iPr₂NEt, DCE, 82%; (j) DDQ, pH 7 buffer, CH₂Cl₂, 92%; (k) TsCl, DABCO, CH₂Cl₂, 90%; (l) allylMgBr, CuI, -20 to 0 °C, 89%. Abbreviations: 9-BBN = 9-borabicyclo-(3.3.1)nonane; TMS = trimethylsilyl; TIP = triisopropylsilyl; MOM = methoxymethyl; DMAP = 4-(dimethylamino)pyridine; NME = *N*methylephedrine; DDQ = 2,3-dichloro-5,6-dicyanobenzoquinone; Ts = tosyl; DABCO = 1,4-diazabicyclo(2.2.2)octane.

Scheme 6. MOM Deprotection Conditions



optimization of this hydroboration reaction with phosphate **5** found the reaction to be highly dependent on the amount of oxidant, equivalents of H₂O, and reaction time. Subsequent PMB ether formation using *p*-methoxybenzyl trichloroacetimidate produced **17** in good yields, demonstrating the acid stability of bicyclic phosphate ($S_1S_1S_2$)-**5**. Employing the previously reported regio- and diastereoselective cuprate addition protocol, displacement of **17** (CuCN · 2LiCl, ZnMe₂, THF, -30 °C to rt) afforded the S_N2' -displaced phosphate acid exclusively (ds \geq 20:1), which upon methylation (TMSCHN₂ and MeOH)

produced cyclic phosphate ester 18 in excellent overall yield (87%). This reaction again highlights the remarkable orbital alignment of the bicyclic phosphate system and its concave nature, where only one of four possible products for this $S_N 2'$ cuprate reaction is generated. Reductive cleavage, followed by sequential protection of the diol systems with TIPS and MOM groups, and final ozonolysis of the olefin afforded aldehyde 19 in good yield.

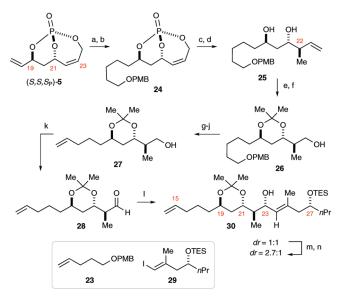
With aldehyde 19 and vinyl iodide 20 in hand, studies aimed at a diastereoselective addition to aldehyde 19 to set the C23 stereogenic carbinol center began (Scheme 5). Initial efforts to generate the C23 stereocenter by lithiate additions gave predominately the undesired 1,2-Felkin product. To overcome this problem, investigation focused on reagent-controlled, ephedrine-based asymmetric vinylzincate additions, described by Marshall.²³ Aldehyde 19 reacted with 20 under these conditions to obtain the desired 1,3-syn isomer 21 in an 11:1 ratio of diastereomers in 65% yield. Successful formation of 21 provided the advance intermediate bearing the requisite stereochemistry of the C15–C30 subunit. With 21 in place, only the installation of the C14-C15 terminal olefin was needed to complete the C15-C30 subunit of dolabelide C. MOM protection of the C23 alcohol, DDQ removal of the PMB ether, tosylation, and cuprate displacement all proceeded in good yield to afford the terminal olefin 22 and complete the C15-C30 subunit of dolabelide C. Overall, a 12-step sequence to 22 from 5 was achieved, bearing the requisite stereochemistry for the C15-C30 subunit of dolabelide C.

IV. Second-Generation Synthesis of the C15–C30 Subunit of Dolabelide C. A second-generation synthesis was next developed when attempts to remove the three MOM-protecting groups from 22 proved problematic. To our dismay, all conditions tested for cleavage of these groups in the presence of the more labile TIPS-protecting groups provided unreacted starting material or total decomposition of the substrate (Scheme 6). The difficulty in removing these protecting groups prompted a reevaluation of protecting groups to access a suitable C15–C30 subunit of dolabelide. This alternative strategy coincided with a planned installation of the C23 carbinol at the last step of the sequence to streamline the route.

The alternative approach to the C15–C30 subunit began by employing previously established CM/reduction methodology (Scheme 7).¹² As anticipated, 5 underwent CM with 23 in the presence of 6 mol % of cat-C¹⁴ (DCE, 90 °C) providing \tilde{E} -configured (>20:1) product in 82% yield. Selective reduction of the external olefin was again achieved using onitrobenzenesulfonyl hydrazine¹⁵ furnishing 24 in 75% yield.²⁴ Compound 24 was also synthesized through a onepot, sequential cross-metathesis/olefin reduction protocol in 59% overall yield utilizing the same reagents shown in Scheme 6. This yield averages to 77% per synthetic step over the two-step combined transformation. Regio- and diastereoselective methyl cuprate addition into 24 and subsequent phosphate cleavage produced diol 25 in good yield.²¹ Diol 25 was protected as the acetonide (PPTS, 2,2-dimethoxypropane) in 96% yield.

The terminal olefin was next converted into primary alcohol **26** by an oxidative cleavage/reduction sequence in good yields. TBS protection of alcohol **26** proceeded in 86% yield and was followed by removal of the PMB ether to provide the corresponding primary alcohol. Conversion of the alcohol to a terminal olefin through an iodination/elimination sequence occurred in excellent yield over the

Scheme 7. Second-Generation Synthesis of C15–C30 Subunit^a



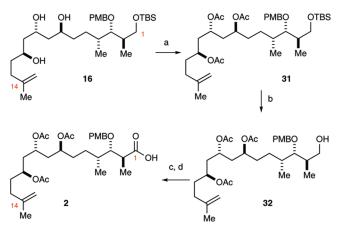
^a Reagents and conditions: (a) cat-C (6 mol %), **23**, DCE, 90 °C, 82%; (b) *o*-NO₂C₆H₅SO₂NHNH₂, Et₃N, CH₂Cl₂, 75%; (c) (1) CuCN·2 LiCl, Me₂Zn, THF, -30 °C to rt; (2) TMSCHN₂, MeOH, 91%; (d) LiAlH₄, Et₂O, 0 °C, 92%; (e) 2,2-DMP, PPTS, CH₂Cl₂, 96%; (f) OsO₄, NMO, *t*-BuOH/THF/H₂O, then NaIO₄, phosphate buffer pH 7, then NaBH₄, EtOH, 0 °C, 81%; (g) TBSCl, pyridine, 95%; (h) H₂, Pd/C, EtOAc, NaHCO₃, 90%; (i) Ph₃P, I₂, imidazole, then *t*-BuOK, THF, 94%; (j) TBAF, THF, 98%; (k) (COCl)₂, DMSO, Et₃N, CH₂Cl₂, -78 °C to rt; (l) *t*-BuLi, Et₂O, **29**, -78 to 0 °C, 30 min, 28, -78 °C, ~1:1 syn:anti, 79% over two steps; (m) Dess-Martin, CH₂Cl₂, 85%; (n) NaBH₄, MeOH, 0 °C, 89%, ~2.7:1 syn:anti. Abbreviations: DMP = dimethoxypropane; PPTS = pyridinium *p*-toluenesulfonate; NMO = *N*-methylmorpholine *N*-oxide; TBS = *tert*-butyldimethylsilyl; TBAF = tetra-*n*-butylammonium fluoride.

two-step sequence. Achievement of the C14/C15 olefin left only a deprotection/oxidation/nucleophilic addition sequence to obtain the necessary C15–C30 subunit. TBAF removal of the TBS-protecting group to **27** and Swern oxidation provided aldehyde **28** necessary for the addition of the C24–C30 fragment.

Despite previous success with the Marshall asymmetric zincate addition protocol,²¹ difficulties in reaction reproducibility and low product yields, also recently noted by Marshall,²³ prompted investigation of an alternative addition sequence. Thus, vinyl iodide **29** was converted to the lithiate with 2 equiv of *t*-BuLi followed by the addition of aldehyde **28** to afford a 1:1 mixture of C23 epimers of alcohol **30** in 79% yield. The two diastereoisomers of **30** were easily separated by column chromatography, allowing for isolation of the correct stereoisomer of alcohol **30** as well as facile recycling (oxidation/reduction) of the undesired diastereomer (dr = 2.7:1).²⁵ Overall, this alternative 13-step route to **30** from phosphate (*S*,*S*,*S*_P)-**5** provided a C15–C30 fragment ready to couple with the C1–C14 subunit of dolabelide C.

V. Completion of the Total Synthesis. Studies toward the completion of dolabelide C next commenced with the complete acetylation of triol **16** to install the proper acetylation pattern for C1–C14 subunit of dolabelide C (Scheme 8). This was accomplished by adding acetic anhydride and pyridine to triol **16** to afford triacetate **31** in excellent yield. Deprotection of the TBS protecting group provided alcohol **32** in 93% yield. Swern

Scheme 8. Generation of Ready-to-Couple C1–C14 Subunit^a



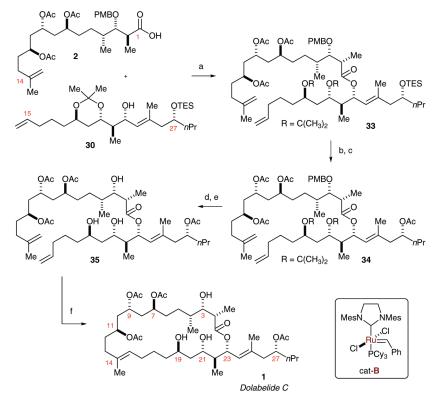
^{*a*} Reagents and conditions: (a) Ac₂O, DMAP, pyridine, 95%; (b) TBAF, THF, 93%; (c) (COCl)₂, DMSO, Et₃N, CH₂Cl₂; (d) NaClO₂, 2-methyl-2-butene, H₃PO₄, 81% over two steps.

oxidation of **32** generated the desired aldehyde that was prone to epimerization and was taken on without purification. Pinnick oxidation of the aldehyde provided carboxylic acid **2** in 81% yield over the two-step sequence, which was ready for coupling with the C15–C30 subunit.

Final coupling of the C1-C14 carboxylic acid 2 and the C15–C30 alcohol 30 was achieved using Yamaguchi conditions²⁶ as previously described by Leighton and co-workers (Scheme 9).⁴ The addition of 2,4,6-trichlorobenzoyl chloride, Et₃N, and DMAP at -78 °C for 21 h avoided epimerization at C2 and yielded the desired coupled 33 in 77% yield. Deprotection of the C27-TES protecting group was achieved with TBAF in 94% yield. Subsequent acylation provided 34 in 98% yield. The final two protecting groups were removed using PPTS in MeOH, followed by treatment with DDQ to provide metathesis precursor 35 in excellent yield over two steps. Efforts to close the ring were attempted prior to PMB ether removal and provided the desired RCM product as observed by HRMS, albeit in poor overall conversion. As a result, subsequent investigations focused on RCM of the deprotected triol 35. Portionwise addition of 20 mol % of (IMesH₂)(PCy₃)(Cl)₂-Ru=CHPh $(cat-B)^{11}$ to triol 35 afforded approximately a 1:1 E/Z mixture of dolabelide C 1 and its (Z)-isomer in a 57-60% yield.

VI. Purification Attempts and RCM/Isomerization Side Reaction. Initial attempts utilizing repeated standard normalphase chromatography removed the Z-isomer (higher R_f), leaving what was believed to be pure dolabelide C(1) as a single spot on TLC. ¹H NMR analysis, however, revealed impurities, which were originally presumed to be the Zisomer. Attempted purification using preparative reversedphase LC-MS revealed the major byproduct, as seen by ${}^{1}H$ NMR, to be a demethylenated analogue (M - 14), along with trace amounts of a constitutional isomer and two byproduct resulting from formal loss of an ethylene (M - 28) as seen in the total-ion chromatogram (Figure 2). These byproducts (35a and 35b) are presumed to occur from isomerization of 35 followed by RCM, resulting in the smaller macrocycles (Scheme 10).²⁷ Numerous reports are consistent with this observation,²⁸ and to the best of our knowledge, this is the first report of a detailed LC-MS analysis of the aforementioned side reaction.

Scheme 9. Completion of Dolabelide C $(1)^a$



^a Reagents and conditions: (a) 2,4,6-trichlorobenzoyl chloride, DMAP, Et₃N, toluene, 77%; (b) TBAF, 94%; (c) Ac₂O, pyridine, DMAP, 98%; (d) PPTS, 83%; (e) DDQ, phosphate buffer pH = 7, CH₂Cl₂, 95%; (f) cat-**B** (20 mol %), CH₂Cl₂ (0.5 mM), 57%, $E:Z = \sim 1:1$.

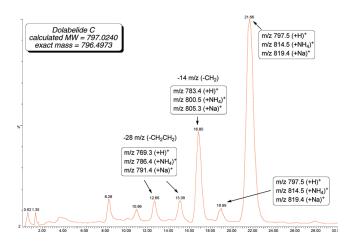


Figure 2. LC–MS analysis of mixture from final RCM.

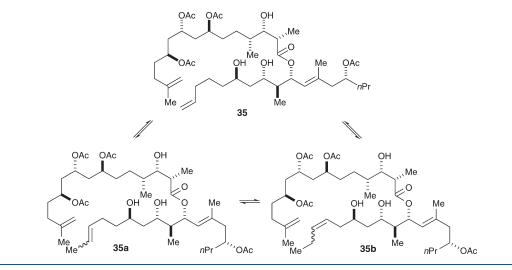
In an attempt to understand and optimize the final RCM sequence, the final metathesis precursor was scaled up. Both syntheses to each subunit proved to be scalable, providing 300 mg of **30** and 175 mg of **32**.²⁹ This material was carried through the same endgame steps (Scheme 9) affording 160 mg of RCM precursor **35** to apply toward the goal of optimizing the C14/15 *E:Z* ratio and minimizing the amount of deleterious side reactions occurring in the final RCM step. For this study, metathesis catalyst cat-**B** and three other candidates shown in Figure 3 were screened.³⁰

Initially, the conditions shown from Scheme 9 were reproduced (Table 1, entry 1), where LC–MS analysis showed nearly complete conversion and a similar ratio to what was previously observed. Screening of other catalysts by varying the phosphine or NHC-ligand showed a significant increase of byproduct formation (entries 3-5). While no significant changes in E/Z ratio were obtained from catalyst screening, rigorous degassing and purification of the solvent were shown to reduce deleterious side products.³¹

Due to the scalability of this synthesis,³² ample material was provided for characterization, where all NMR spectra matched with the previous reported data.^{2,33} In addition, sufficient quantities of the unnatural C14/15 Z-isomer³⁴ were generated for both NMR analysis and collection of biological data to determine its bioactivity and potential potency against cervical cancer.³⁵

In conclusion, dolabelide C (1) and its non-natural C14–C15 Z-diastereomer were produced and isolated from a scalable phosphate-mediated synthesis. A complex mixture was generated in the final RCM step resulting in byproduct, which arose from a net loss of CH₂ and C₂H₄, that proved to be difficult to separate via repeated flash chromatography (8:1 CH₂Cl₂:acetone). Since the material produced at the end of the first synthesis of 1 was sparse, a resynthesis provided 175 mg of the C1–C14 subunit (32), 300 mg of the C15–C30 subunit (30), and 160 mg of RCM precursor 35. This allowed a detailed optimization study through a screening of various metathesis catalysts, concluding that the originally developed conditions (20 mol % cat-B, 0.5 μ m, 40 °C) provided the optimum results. Overall, 14 mg of pure dolabelide C and 10 mg of the pure Z-isomer were produced in a 24-step longest linear

Scheme 10. Possible Isomerization Pathways from Metathesis Step



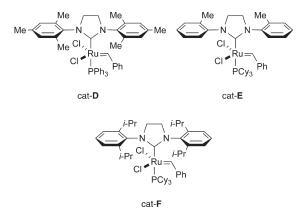


Figure 3. Metathesis catalysts screened on final RCM step.

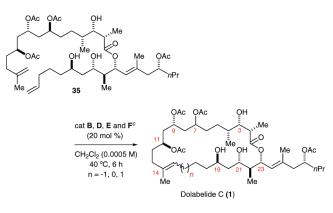
sequence (LLS) from commercially available material utilizing the (R_r,R_r,R_p) antipode of **5** (sequence streamlined to a 22-step LLS using one-pot, sequential protocols).

EXPERIMENTAL SECTION

General Methods. All reactions were carried out in oven- or flamedried glassware, under an argon atmosphere, using standard gastight syringes, cannulae, and septa. Stirring was achieved with oven-dried magnetic stir bars. Et₂O, THF, and CH₂Cl₂ were passed through a purification system employing activated Al₂O₃. Et₃N was eluted through basic alumina and stored over KOH. Butyllithium was titrated prior to use. All olefin metathesis catalysts were obtained commercially and used without further purification. All ¹H and ¹³C NMR spectra were recorded in CDCl₃ or C₃D₅N at 400 or 500 MHz and 126 MHz, respectively, and calibrated to the solvent peak. All mass spectra were obtained using electrospray ionization (ESI) (MeOH) coupled to high-resolution mass spectrometry (HRMS). Observed rotations at 589 nm were measured using an automatic polarimeter. Infrared spectra were obtained using a Fourier-transform infrared (FTIR) spectrometer.

(45,6*R*)-4-((*R*)-But-3-en-2-yl)-6-(5-(4-methoxybenzyloxy)pentyl)-2,2-dimethyl-1,3-dioxane (Sl-1). Diol 25 (2.00 g, 5.95 mmol) was dissolved in CH_2Cl_2 (8 mL) at rt. 2,2-Dimethoxypropane

Table 1. Catalyst Screening of Final RCM



entry	catalyst	conversion (%)	E/Z^a	<i>E</i> / <i>Z</i> /byproducts ^{<i>a</i>}
1	cat-B	>99	1:1	1:1:0.17
2	$cat-B^b$	87	1.2:1	1.2:1:0.26
3	cat-D	100	1:1	1:1:0.80
4	cat-E	100	1:1.1	1:1.1:0.45
5	cat-F	87	1.2:1	1.2:1:0.61

^{*a*} Ratios determined through peak area integration from LC–MS analysis of crude mixtures. ^{*b*} Purified newly purchased catalyst through SiO₂ plug in 10:1 hexanes/EtOAc. ^{*c*} All reactions were run with stepwise addition of 20 mol % of catalyst over 6 h at 40 °C in 0.5 μ M CH₂Cl₂.

(8 mL) and PPTS (150 mg, 0.595 mmol) were added, respectively, and the clear solution was stirred until completion. The reaction was quenched with saturated NaHCO₃ (15 mL) and diluted with CH₂Cl₂. The aqueous layer was extracted with CH₂Cl₂ (3 × 25 mL), and the combined organic layers were washed once with brine (30 mL), dried (Na₂SO₄), and concentrated under reduced pressure. Compound **SI-1** was isolated using flash chromatography (19:1 hexanes/EtOAc) as a clear oil (2.18 g, 98%): [α]_D = -36.3 (*c* = 0.40, CH₂Cl₂); FTIR (neat) 2983, 2935, 2856, 1612, 1512, 819 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 7.25 (d, *J* = 8.4 Hz, 2H), 6.87 (d, *J* = 8.6 Hz, 2H), 5.83 (ddd, *J* = 17.4, 10.5, 7.3 Hz, 1H), 5.03 (ddd, *J* = 17.5, 11.0, 2.6 Hz, 2H), 4.43 (s, 2H), 3.80 (s, 3H), 3.74–3.66 (m, 1H), 3.63 (ddd, *J* = 9.7, 6.3, 6.3 Hz, 1H),

3.45 (t, J = 6.6 Hz, 2H), 2.18–2.26 (m, 1H), 1.71–1.64 (m, 1H), 1.63–1.53 (m, 2H), 1.55–1.35 (m, 6H), 1.32 (s, 6H), 1.30–1.24 (m, 1H), 0.98 (d, J = 6.9 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 159.1, 140.9, 130.7, 129.3, 114.4, 113.7, 100.3, 72.5, 70.1, 70.0, 66.8, 55.3, 42.1, 36.3, 35.9, 29.7, 26.2, 25.3, 24.7, 24.4, 15.3; HRMS calcd for C₂₃H₃₆NaO₄ (M + Na)⁺ 399.2511, found 399.2498 (ESI).

(R)-2-((4S,6R)-6-(5-(4-Methoxybenzyloxy)pentyl)-2,2-dimethyl-1,3-dioxan-4-yl)propan-1-ol (26). Olefin SI-1 (1.50 g, 3.99 mmol) was dissolved in t-BuOH/THF/H₂O (10:2:1, 20 mL) at rt. N-Methylmorpholine N-oxide (933 mg, 7.98 mmol) and OsO4 (0.19 mL, 0.08 mmol, 4% aq) were added, and the reaction was stirred for approximately 12 h until olefin was completely consumed. The mixture was then diluted with phosphate buffer pH 7 (twice the volume of *t*-BuOH), and NaIO₄ was added (3.41 mg, 16.0 mmol). The reaction was stirred vigorously for approximately 2 h until the diol was completely consumed (monitored by TLC). The reaction was quenched with solid Na_2SO_3 (2.0 g), and acetone was removed under reduced pressure. The residue was partitioned with EtOAc (20 mL) and H₂O (10 mL), and the aqueous layer was extracted with EtOAc (3 \times 20 mL). The collected organics were washed once with brine (20 mL), dried (Na₂SO₄), and filtered. After concentration under reduced pressure, the crude product was purified using flash chromatography (5:1 hexanes/EtOAc) to generate intermediate aldehyde as a yellow oil.

The resultant aldehyde was dissolved in EtOH (16 mL) and cooled to 0 °C. NaBH₄ (303 mg, 7.98 mmol) was added, and the reaction was slowly brought back to rt. Upon completion (\sim 45 min), the solution was partitioned with 2:1 Et₂O/H₂O (40 mL), the aqueous layer was extracted with Et₂O (3×5 mL), and the organic layers were combined, washed with brine (30 mL), dried (MgSO₄), filtered, and concentrated under reduced pressure. Purification with flash chromatography (1:2 hexanes/EtOAc) afforded 26 (88% over two steps, 1.35 mg) as a clear oil: $[\alpha]_D = -0.26$ (*c* = 0.18, CH₂Cl₂); FTIR (neat) 3442, 2933, 2856, 1612, 1512, 819 cm $^{-1}$; ¹H NMR (500 MHz, CDCl₃) δ 7.25 (d, J = 8.5 Hz, 2H), 6.87 (d, J = 8.6 Hz, 2H), 4.43 (s, 2H), 3.81 (s, 3H), 3.79–3.73 (m, 1H), 3.68 (ddd, J = 9.2, 6.3, and 6.3 Hz, 1H), 3.58 (d, J = 5.1 Hz, 2H), 3.43 (t, J = 6.6 Hz, 2H), 3.08 (s, 1H), 1.20–1.80 (m, 11H), 1.38 (s, 3H), 1.33 (s, 3H), 0.82 (d, J = 7.0 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 159.1, 130.8, 129.2, 113.7, 100.5, 73.1, 72.5, 70.1, 68.3, 66.6, 55.3, 40.6, 37.9, 35.8, 29.7, 26.2, 25.2, 24.6, 24.6, 12.7; HRMS calcd for $C_{22}H_{36}NaO_5 (M + Na)^+$ 403.2460, found 403.2413 (ESI).

tert-Butyl((R)-2-((4S,6R)-6-(5-(4-methoxybenzyloxy)pentyl)-2,2-dimethyl-1,3-dioxan-4-yl)propoxy)dimethylsilane (SI-2). Alcohol 26 (1.34 mg, 3.53 mmol) was dissolved in CH2Cl2 (23 mL) at rt. Imidazole (720 mg, 10.6 mmol), DMAP (10 mg, 0.08 mmol), and TBSCl (800 mg, 5.29 mmol) were added, respectively. The reaction was quenched upon completion (~90 min, monitored by TLC) with saturated NH₄Cl (25 mL) and diluted with Et₂O (50 mL). The aqueous layer was extracted with Et₂O (3 \times 25 mL), and the organic layers were washed with brine (25 mL). The combined organic layers were dried (Na₂SO₄) and concentrated under reduced pressure. Purification with flash chromatography (20:1 hexanes/EtOAc) afforded **SI-2** (1.70 g, 97%) as a yellow oil: $[\alpha]_D = -16.6$ (c = 0.35, CH₂Cl₂); FTIR (neat) 2933, 2856, 2881, 1247, 835 cm⁻¹; ¹H NMR (500 MHz, $CDCl_3$) δ 7.26 (d, J = 8.4 Hz, 2H), 6.88 (d, J = 8.6 Hz, 2H), 4.24 (s, 2H), 3.80(s, 3H), 3.74-3.66(m, 2H), 3.56(d, J = 4.4 Hz, 2H), 3.44(t, J = 6.6 Hz, 3.80(s, 3H))2H), 1.37–1.67 (m, 11H), 1.31 (s, 6H), 0.89 (s, 9H), 0.85 (d, J = 6.8 Hz, 3H), 0.02 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 159.0, 130.7, 129.2, 113.7, 100.1, 72.5, 70.1, 67.0, 66.8, 64.1, 55.2, 40.5, 36.6, 35.9, 29.7, 26.1, 25.9, 25.3, 24.6, 24.5, 18.2, 12.2, -5.5, -5.5; HRMS calcd for $C_{28}H_{50}NaO_5Si (M + Na)^+$ 517.3325, found 517.3334 (ESI).

5-((4R,6S)-6-((R)-1-(tert-Butyldimethylsilyloxy)propan-2yl)-2,2-dimethyl-1,3-dioxan-4-yl)pentan-1-ol (SI-3). PMB ether SI-2 (1.65 g, 3.34 mmol) was dissolved in EtOAc (16 mL) at rt. A catalytic amount of 10% Pd/C (50 mg) and NaHCO₃ (280 mg, 3.34 mmol) were added sequentially, and the flask was pressurized with a H₂ balloon. After 10 h, the mixture was filtered through pad of Celite and rinsed thoroughly with EtOAc. The filtrate was concentrated under reduced pressure and purified with flash chromatography (10:1 hexanes/EtOAc) to yield alcohol **SI-3** (1.12 g, 90% yield) as a clear oil: $[\alpha]_D = -0.11$ (c = 0.40, CH₂Cl₂); FTIR (neat) 3357, 2933, 2858, 1379, 1251, 1224, 835, 775 cm⁻¹; ¹H NMR (500 MHz, CDCl₃)^{36a} δ 3.75–3.68 (m, 2H), 3.63 (t, J = 6.6 Hz, 2H), 3.55 (dd, J = 4.7 and 1.3 Hz, 2H), 1.70–1.38 (m, 10H), 1.33 (s, 6H), 1.28–1.22 (m, 1H), 0.89 (s, 9H), 0.84 (d, J = 6.9 Hz, 3H), 0.04 (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 100.2, 67.1, 66.8, 64.1, 63.0, 40.5, 36.6, 35.9, 32.7, 25.9 (3), 25.7, 25.3, 24.6, 24.5, 18.2, 12.2, -5.5, -5.5; HRMS calcd for C₂₀H₄₂NaO₄Si (M + Na)⁺ 397.2750, found 397.2773 (ESI).

(*R*)-2-((45,6*R*)-2,2-Dimethyl-6-(pent-4-enyl)-1,3-dioxan-4yl)propan-1-ol (27). Alcohol SI-3 (1.12 g, 2.99 mmol) was dissolved in THF (30 mL) at rt. Triphenylphosphine (941 mg, 3.59 mmol) and imidazole (477 mg, 6.59 mmol) were added, respectively, and the solution was cooled to 0 °C. I₂ (912 mg, 3.59 mmol) was added, and the reaction was stirred for approximately 30 min (monitored by TLC). The solution was diluted with hexane and filtered through a pad of silica, while washing with hexane, and concentrated under reduced pressure. The crude product was taken onto the next step.

The iodo compound was dissolved in THF (35 mL) at rt followed by stepwise addition of *t*-BuOK (1.0 g, 8.98 mmol). The reaction was stirred for \sim 30 min and was quenched with H₂O. The aqueous layer was extracted with EtOAc (3 × 20 mL portions), and the organic layers were combined, washed with brine (20 mL), dried (Na₂SO₄), filtered, and concentrated under reduced pressure. Purification with flash chromatography (20:1 hexanes/EtOAc) yielded the resultant terminal olefin (1.0 g, 94%) as a clear oil.

The resultant silyl ether (1.00 g, 2.81 mmol) was dissolved in THF (10 mL) and cooled to 0 °C. A solution of TBAF in THF (8.5 mL, 1.0 M in THF) was added dropwise. The reaction was stirred at 0 °C until completion (\sim 45 min) and quenched with saturated NH₄Cl (10 mL), and the aqueous layer was extracted with Et₂O (3 \times 20 mL). The organic layers were combined, washed with brine (20 mL), dried (Na₂-SO₄), filtered, and concentrated under reduced pressure. Purification using flash chromatography (10:1 hexanes/EtOAc) afforded 27 (670 mg, 98%) as a clear oil: $[\alpha]_{D} = -78.6$ (*c* = 0.50, CH₂Cl₂); FTIR (neat) 3446, 2983, 2935, 2879, 1379, 1224, 908 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 5.86-5.75 (m, 1H), 5.07-4.93 (m, 2H), 3.82-3.76 (m, 1H), 3.69 (ddd, I = 9.2, 9.2, and 6.2 Hz, 1H), 3.61 - 3.55 (m, 2H), 3.08 (s, 1H),2.06 (dd, I = 14.1 and 7.1 Hz, 2H), 1.80–1.40 (m, 7H), 1.38 (s, 3H), 1.33 (s, 3H), 0.81 (d, J = 7.0 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 138.7, 114.7, 100.5, 73.0, 68.2, 66.6, 40.6, 37.8, 35.3, 33.6, 24.7, 24.6, 24.6, 12.6; HRMS calcd for $C_{14}H_{27}O_3$ (M + H)⁺ 243.1960, found 243.2895 (ESI).

(2R,3R,7R,E)-2-((4S,6R)-2,2-Dimethyl-6-(pent-4-enyl)-1,3dioxan-4-yl)-5-methyl-7-(triethylsilyloxy)dec-4-en-3-ol (30). A solution of oxalyl chloride (0.158 mL, 1.86 mmol) in CH₂Cl₂ (4.8 mL) was cooled to -78 °C, and DMSO (0.220 mL, 3.01 mmol) was added slowly by syringe (gas evolution). After being stirred for 10 min, a solution of alcohol 27 (300 mg, 1.24 mmol) in CH₂Cl₂ (3.0 mL) was added by cannula and rinsed with CH_2Cl_2 (2 × 0.5 mL). The cloudy mixture was stirred at -78 °C for 15 min at which time Et₃N (0.700 mL, 4.96 mmol) was added dropwise. The reaction mixture was stirred for 1 h at -78 °C, quenched cold with saturated NaHCO₃ (5 mL), and allowed to warm to rt. After dilution with CH2Cl2, the layers were separated, and the aqueous layer was re-extracted with CH_2Cl_2 (3 × 12 mL). The organic layer was dried (Na₂SO₄), filtered through a silica plug, and rinsed (3 \times 25 mL) with EtOAc/CH₂Cl₂(1:1). The filtrate was concentrated under reduced pressure to give aldehyde 28 as a yellow oil. The crude aldehyde was taken immediately to the next reaction without further purification.

To a solution of the vinyl iodide **29** (1.01 mg, 2.75 mmol) in Et₂O (10 mL) at -78 °C was added *t*-BuLi (1.7 M in pentane, 3.40 mL, 5.75 mmol), and the reaction was immediately warmed to 0 °C for 25 min. The reaction was recooled to -78 °C, and the aldehyde **28** was slowly added via syringe in Et₂O (2.5 mL, 0.60 mL rinse). After 1 h, the reaction was quenched at -78 °C with saturated NH₄Cl and warmed to rt, and the layers were separated. The aqueous layer was extracted with Et₂O (2 × 10 mL), and the combined organic layers washed with brine (15 mL), dried (Na₂SO₄), filtered, and concentrated under reduced pressure. Flash chromatography (10:1 hexanes/EtOAc) afforded a 1:1 mixture of 1,3-*syn* and 1,3-*anti* **30** (ratio determined by ¹H NMR analysis of crude reaction mixture, 464 mg, combined yield of diastereomers 77%

over two steps). **Oxidation/Reduction Sequence.** The 1,3-*anti* diastereomer (65 mg, 0.135 mmol) of **30** was dissolved in CH₂Cl₂ (2.6 mL) at rt. Dess-Martin periodinane (115 mg, 0.270 mmol) was added to the stirring solution, where upon completion (monitored by TLC) the reaction was diluted with Et₂O (5 mL). The organic layer was washed with saturated NaHCO₃ (2 × 5 mL) and dried (Na₂SO₄). After filtration, the solvent was removed under reduced pressure, and the residual oil was purified through a short plug of SiO₂ (1:1 hexanes/ EtOAc) to provide a clear oil (40 mg, 85%).

The ketone (6 mg, 0.0125 mmol) was dissolved in MeOH and cooled to 0 °C. NaBH₄ (11 mg, 0.035 mmol) was added slowly, and the mixture was stirred until the ketone was completely consumed (monitored by TLC). The mixture was partitioned with H₂O/Et₂O (1:1, 10 mL), and the resultant aqueous layer was extracted with Et₂O (3×5 mL). The collected organic layers were washed with brine (5 mL) and dried (Na₂SO₄). The epimeric ratio of the crude material was determined by ¹H NMR analysis after filtration and removal of solvent under reduced pressure (~2.7:1). Flash chromatography (5:1 hexanes/EtOAc) provided both isomers (4 mg, 89%) as a clear oil: $[\alpha]_D = -8.1$ (c = 1.3, CH₂Cl₂); FTIR (neat) 3456, 2954, 2935, 2875, 1458, 1379, 1224, 908 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 5.82 (dddd, J = 16.9, 10.1, 6.7, 6.7 Hz, 1H), 5.16 (d, J = 9.1 Hz, 1H), 5.01 (ddd, J = 17.1, 3.4, 1.5 Hz, 1H), 4.95 (d, J = 10.2 Hz, 1H), 4.27 (t, J = 8.7 Hz, 1H), 3.99 (s, 1H), 3.84–3.74 (m, 3H), 2.27 (dd, J = 14.1, 4.3 Hz, 1H), 2.16 (dd, J = 8.7, 3.0 Hz, 1H), 2.06 (q, J = 7.0 Hz, 2H), 1.70 (s, 3H), 1.41 (s, 3H), 1.35 (s, 3H), 1.72 - 1.20 (m, 11H), 0.96 (t, J = 8.2 Hz, 9H), 0.88 (t, J = 7.3 Hz, 3H), 0.71 (d, J = 6.9 Hz, 3H), 0.59 (q, J = 8.2 Hz, 6H); ¹³C NMR (126 MHz, CDCl₃) & 138.6, 136.0, 128.8, 114.6, 100.6, 72.9, 72.4, 70.7, 66.6, 48.5, 44.1, 38.7, 38.0, 35.2, 33.6, 24.7, 24.6, 24.5, 18.4, 17.3, 14.2, 11.6, 7.0, 5.0; HRMS calcd for $C_{28}H_{54}NaO_4Si (M + Na)^+$ 505.3689, found 505.3674 (ESI).

(5S,7R,9S,12R,13S,14R)-15-(tert-Butyldimethylsilyloxy)-13-(4-methoxybenzyloxy)-2,12,14-trimethylpentadec-1-ene-5,7,9-triyl Triacetate (31). To a solution of triol 16 (132 mg, 0.232 mmol) in CH₂Cl₂ (3.3 mL) were added DMAP (3 mg, 0.023 mmol), pyridine (0.750 mL, 9.30 mmol), and acetic anhydride (0.45 mL, 4.65 mmol). The reaction was stirred until the disappearance of starting material at rt (\sim 2 h). The reaction was diluted with EtOAc (5 mL) and quenched with saturated NH₄Cl (5 mL), and the aqueous layer was reextracted with EtOAc (3 \times 10 mL). The organic layer was washed with brine (10 mL), dried (Na₂SO₄), filtered, and concentrated under reduced pressure. Flash chromatography (5:1 hexanes/EtOAc) provided 31 (151 mg, 94%) as a clear oil: $[\alpha]_{\rm D} = +12.4$ (*c* = 0.50, CH₂Cl₂); FTIR (neat) 2956, 2929, 2883, 2856, 1739, 1514, 1461, 1247 cm⁻¹; ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3) \delta$ 7.26 (d, J = 8.7 Hz, 2H), 6.87 (d, J = 8.7 Hz, 2H), 4.97 (dddd, J = 9.5, 6.2, 6.2, 3.1 Hz, 1H), 4.89 (m, 2H), 4.71 (s, 1H), 4.66 (s, 1H), 4.53 (d, J = 10.9 Hz, 1H), 4.46 (d, J = 10.9 Hz, 1H), 3.80 (s, 3H), 3.71 (dd, J = 9.7, 5.3 Hz, 1H), 3.63 (dd, J = 9.7, 3.3 Hz, 1H), 3.25 (dd, J = 8.7, 2.4 Hz, 1H), 2.07–1.99 (m, 2H), 2.05 (s, 3H), 2.03 (s, 3H), 1.99 (s, 3H), 1.92 (dddd, J = 18.7, 10.2, 4.4, 4.4 Hz, 1H), 1.84-1.68 (m, 5H), 1.71 (s, 3H), 1.64-1.54 (m, 4H), 1.46-1.38 (m, 1H), 1.34–1.22 (m, 1H), 0.92 (s, 9H), 0.90 (d, J = 2.3 Hz, 3H), 0.88 (d, J =

2.3 Hz, 3H), 0.06 (s, 6H); 13 C NMR (126 MHz, CDCl₃) δ 170.8, 170.7, 170.6, 159.1, 144.9, 131.7, 129.3, 113.9, 110.5, 83.3, 74.6, 70.9, 70.3, 67.5, 65.1, 55.4, 39.2, 38.7, 38.6, 35.4, 33.5, 33.1, 32.3, 30.5, 26.1, 22.6, 21.3, 21.3, 21.2, 18.5, 14.8, 13.6, -5.2, -5.2; HRMS calcd for C₃₈H₆₄NaO₉Si (M + Na)⁺ 715.4217, found 715.4213 (ESI).

(5S,7R,9S,12R,13S,14R)-15-Hydroxy-13-(4-methoxybenzyloxy)-2,12,14-trimethylpentadec-1-ene-5,7,9-triyl Triacetate (32). To a solution of 31 (150 mg, 0.216 mmol) in THF (2.3 mL) was added TBAF (0.70 mL, 1.0 M in THF). The reaction was stirred until disappearance of starting material at rt (\sim 3 h). The reaction was diluted with EtOAc (3 mL) and guenched with saturated NH₄Cl (5 mL), and the aqueous layer was re-extracted with EtOAc (2 \times 5 mL). The organic layer was dried (Na₂SO₄), filtered, and concentrated under reduced pressure. Flash chromatography (2:1 hexanes/EtOAc) provided 32 (118 mg, 94%) as a clear oil: $[\alpha]_D = +13.1$ (c = 2.4, CH₂Cl₂); FTIR (neat) 3502, 3072, 2964, 2935, 2875, 1737, 1514, 1454, 1245 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.28 (d, J = 8.5 Hz, 2H), 6.88 (d, J = 8.6 Hz, 2H), 4.98 (dddd, J = 9.6, 6.3, 6.3, 3.3 Hz, 1H), 4.99-4.85 (m, 2H), 4.73 (s, 1H), 4.67 (s, 1H), 4.58 (d, J = 10.6 Hz, 1H), 4.51 (d, J = 10.6 Hz, 1H), 3.81 (s, 3H), 3.62–3.67 (m, 2H), 3.25 (dd, J = 7.6, 3.3 Hz, 1H), 2.71 (s, 1H), 2.07-1.99 (m, 2H), 2.06 (s, 3H), 2.04 (s, 3H), 2.02 (s, 3H), 1.95–1.86 (m, 2H) 1.84–1.68 (m, 6H), 1.72 (s, 3H), 1.64–1.56 (m, 2H), 1.54–1.44 (m, 1H), 1.34–1.22 (m, 1H), 0.96 (d, J = 6.9 Hz, 3H), 0.94 (d, J = 7.1 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 170.7, 170.6, 170.5, 159.3, 144.7, 130.4, 129.4 (2), 113.9 (2), 110.4, 87.6, 74.8, 70.6, 70.0, 67.3, 66.5, 55.3, 39.0, 38.4, 37.7, 36.1, 33.3, 32.8, 32.2, 30.0, 22.4, 21.2, 21.1, 21.1, 15.4, 14.3; HRMS calcd for C₃₂H₅₀NaO₉ (M $+ Na)^+$ 601.3353, found 601.3354 (ESI).

(2S,3S,4R,7S,9R,11S)-7,9,11-Triacetoxy-3-(4-methoxybenzyloxy)-2,4,14-trimethylpentadec-14-enoic Acid (2). A solution of oxalyl chloride (0.046 mL, 0.539 mmol) in CH₂Cl₂ (1.67 mL) was cooled to -78 °C, and DMSO (0.077 mL, 1.08 mmol) was added slowly by syringe (gas evolution). After the mixture was stirred for 10 min, a solution of alcohol 32 (125 mg, 0.21 mmol) in CH₂Cl₂ (2.0 mL) was added by cannula and rinsed with CH_2Cl_2 (2 × 0.2 mL). The cloudy mixture was stirred at -78 °C for 15 min, at which time Et₃N (0.18 mL, 1.29 mmol) was added dropwise. The reaction mixture was stirred for 2 h at -78 °C. The reaction was guenched at -78 °C with saturated NaHCO₃ (3 mL) and allowed to warm to rt. The reaction was diluted with CH₂Cl₂, and the layers were separated. The aqueous layer was reextracted with CH_2Cl_2 (3 \times 5 mL). The organic layer was dried (Na₂SO₄), filtered, and concentrated under reduced pressure to give aldehyde as a yellow oil. The crude aldehyde was taken immediately to the next reaction with further purification.

To a solution of crude aldehyde were added tert-butyl alcohol (4.5 mL) and 2-methyl-2-butene (1.5 mL). A solution of NaClO2 (390 mg, 4.30 mmol) and sodium dihydrogen phosphate (470 mg, 3.01 mmol) in H₂O (2.0 mL) was prepared and added to the reaction mixture by syringe. The yellow solution was stirred vigorously for 2 h at rt, diluted with Et₂O (15 mL), and poured into H₂O (9 mL). The layers were separated, and the aqueous phase was extracted with Et₂O (3 \times 10 mL). The combine organic layers were dried (Na₂SO₄), filtered, and concentrated under reduced pressure. Flash chromatography (1:1 hexanes/EtOAc) provided 2 (104 mg, 81% over two steps) as a clear oil: $[\alpha]_{D} = +8.13$ (*c* = 0.16, CH₂Cl₂); FTIR (neat) 3251, 3076, 2964, 2923, 2854, 1737, 1714, 1512, 1454, 1245 cm⁻¹; ¹H NMR (400 MHz, $CDCl_3$) δ 7.24 (d, J = 8.5 Hz, 2H), 6.85 (d, J = 8.6 Hz, 2H), 5.10-4.85 (m, 3H), 4.73 (s, 1H), 4.67 (s, 1H), 4.56 (d, J = 10.7 Hz, 1H), 4.50 (d, J = 10.6 Hz, 1H), 3.79 (s, 3H), 3.57 (dd, J = 7.2, 3.5 Hz, 1H), 2.78 (dddd, J = 14.33, 7.1, 7.1, 7.1 Hz, 1H), 2.07-1.99 (m, 2H), 2.06 (s, 3H), 2.04 (s, 3H), 2.02 (s, 3H), 1.95–1.86 (m, 2H), 1.84–1.64 (m, 5H), 1.72 (s, 3H), 1.64–1.40 (m, 3H), 1.39–1.20 (m, 2H), 1.17 (d, J = 7.1 Hz, 3H), 0.93 $(d, J = 6.7 \text{ Hz}, 3\text{H}); {}^{13}\text{C} \text{ NMR} (126 \text{ MHz}, \text{CDCl}_3) \delta 179.9, 170.8, 170.7,$ 170.5, 159.4, 144.7, 130.0, 129.5 (2), 113.9 (2), 110.3, 84.2, 74.4, 70.8, 69.9, 67.3, 55.3, 42.3, 39.0, 38.1, 35.6, 33.3, 32.5, 32.3, 28.9, 22.4, 21.2, 21.1, 21.1, 14.7, 14.4; HRMS calcd for $C_{32}H_{48}NaO_{10}$ (M + Na)⁺ 615.3145, found 615.3131 (ESI).

(5S,7R,9S,12R,13S,14S)-15-((2S,3R,7R,E)-2-((4S,6R)-2,2-Dimethyl-6-(pent-4-enyl)-1,3-dioxan-4-yl)-5-methyl-7-(triethylsilyloxy)dec-4-en-3-yloxy)-13-(4-methoxybenzyloxy)-2,12, 14-trimethyl-15-oxopentadec-1-ene-5,7,9-triyl Triacetate (33). To a solution of alcohol 30 (77 mg, 0.159 mmol), carboxylic acid 2 (106 mg, 0.175 mmol), and DMAP (975 mg, 7.98 mmol) in toluene (32 mL) at -78 °C was added Et₃N (0.5 mL, 3.61 mmol) dropwise followed by the slow addition of 2,4,6-trichlorobenzoyl chloride (0.56 mL, 3.58 mmol), which caused the white solution to thicken. The mixture was stirred for 21 h at -78 °C ensuring that the bath temperature did not rise above -65 °C. The reaction flask was then moved to a dry ice/CH₃CN bath and stirred for 2.5 h maintaining the temperature between -30 and -42 °C. At the end of the 2.5 h, the solution was slowly allowed to warm to rt in the bath over 1 h. The flask was placed in an ice bath for 2 h while being stirred. The reaction was quenched by the addition of saturated NaHCO₃ (15 mL). The layers were separated, and the aqueous layer was back-extracted with Et₂O (25 mL). The combined organic layers were dried (Na₂SO₄), filtered, and concentrated under reduced pressure. Purification by flash chromatography (5:1 hexanes/EtOAc) provided ester 33 (130 mg, 77%) as a colorless oil: $[\alpha]_{\rm D}$ +3.63 (c = 0.28, CH₂Cl₂); FTIR (neat) 3076, 2954, 2935, 2875, 1739, 1515, 1442, 1244 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.19 (d, J = 8.7 Hz, 2H), 6.83 (d, J = 8.7 Hz, 2H), 5.82 (dddd, J = 17.0, 10.2, 6.8, 6.8 Hz, 1H), 5.68 (dd, *J* = 9.9, 5.7 Hz, 1H), 5.17 (d, *J* = 9.8 Hz, 1H), 5.02 (ddd, *J* = 17.1, 3.2, 1.6 Hz, 1H), 5.00-4.93 (m, 3H), 4.94-4.85 (m, 2H), 4.74 (s, 1H), 4.67 (s, 1H), 4.51 (d, J = 10.8 Hz, 1H), 4.34 (d, J = 10.8 Hz, 1H), 3.79 (s, 3H), 3.70-3.80 (m, 2H), 3.69-3.60 (m, 3H), 2.69 (dt, J = 14.2, 7.1 Hz, 1H), 2.20–1.89 (m, 8H), 2.06 (s, 3H), 2.04 (s, 3H), 2.00 (s, 3H), 1.80–1.20 (m, 18H), 1.76 (s, 3H), 1.73 (s, 3H), 1.34 (s, 3H), 1.29 (s, 3H), 1.08 (d, J = 7.1 Hz, 3H), 0.96 (t, J = 7.9 Hz, 9H), 0.93-0.84 (m, 9H) 0.58 (q, J = 7.9 Hz, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 175.3, 170.6, 170.6, 170.5, 158.9, 144.7, 139.6, 138.8, 131.2, 129.0, 122.7, 114.6, 113.6, 110.3, 100.1, 83.1, 73.9, 71.4, 70.7, 70.2, 70.0, 67.3, 67.1, 66.5, 55.2, 53.5, 48.8, 43.5, 42.1, 39.1, 38.5, 38.5, 35.4, 34.7, 33.7, 33.3, 32.1, 30.3, 29.9, 29.7, 24.9, 24.9, 24.8, 22.4, 21.2, 21.1, 21.1, 18.3, 17.6, 15.3, 14.8, 14.2, 13.2, 9.8, 7.0 (3), 5.0 (3); HRMS calcd for $C_{60}H_{100}NaO_{13}Si (M + Na)^+$ 1079.6831, found 1079.7115 (ESI).

(5S,7R,9S,12R,13S,14S)-15-((2S,3R,7R,E)-2-((4S,6R)-2,2-Dimethyl-6-(pent-4-enyl)-1,3-dioxan-4-yl)-7-hydroxy-5-methyldec-4-en-3-yloxy)-13-(4-methoxybenzyloxy)-2,12,14-trimethyl-15-oxopentadec-1-ene-5,7,9-triyl Triacetate (SI-4). Ester 33 (75 mg, 0.071 mmol) was dissolved in THF (1 mL) and cooled to 0 °C. A solution of TBAF in THF (0.214 mL, 1.0 M in THF) was added dropwise. The reaction stirred at 0 °C until completion (approximately 45 min). The reaction was quenched with saturated NH₄Cl, and the aqueous layer was extracted with EtOAc (3 \times 10 mL). The organic layers were combined, washed with brine (5 mL), dried (Na₂SO₄), filtered, and concentrated under reduced pressure. Purification by flash chromatography (2:1 hexanes/EtOAc) afforded SI-4 (63 mg, 94%) as a clear oil: $[\alpha]_{\rm D} = -11.4 (c = 1.0, \text{CHCl}_3); {}^{1}\text{H NMR} (400 \text{ MHz}, \text{CDCl}_3) \delta 7.22 (d,$ *J* = 8.7 Hz, 2H), 6.84 (d, *J* = 8.7 Hz, 2H), 5.81 (dddd, *J* = 17.0, 10.2, 6.7, and 6.7 Hz, 1H), 5.64 (dd, J = 9.9 and 5.2 Hz, 1H), 5.20 (d, J = 9.6 Hz, 1H), 5.01 (ddd, J = 17.1, 3.4, and 1.6 Hz, 1H), 5.00–4.93 (m, 2H), 4.93–4.85 (m, 2H), 4.72 (s, 1H), 4.67 (s, 1H), 4.53 (d, J = 10.7 Hz, 1H), 4.38 (d, J = 10.7 Hz, 1H), 3.79 (s, 3H), 3.76-3.69 (m, 1H), 3.66-3.59 (m, 1H), 3.60–3.54 (m, 1H), 3.53 (dd, J = 8.4, 3.0 Hz, 1H), 2.70 (dt, J = 15.2 and 7.2 Hz, 1H), 2.15–1.87 (m, 8H), 2.06 (s, 3H), 2.04 (s, 3H), 2.00 (s, 3H), 1.80–1.20 (m, 21H), 1.76 (s, 3H), 1.73 (s, 3H), 1.33 (s, 3H), 1.28 (s, 3H), $1.10 (d, J = 7.1 Hz, 3H), 0.93 - 0.84 (m, 9H); {}^{13}C NMR (126 MHz, CDCl_3)$ δ 175.1, 170.7, 170.6, 170.5, 158.9, 144.7, 139.6, 138.7, 131.2, 128.8, 123.1, 114.6, 113.5, 110.3, 100.2, 83.7, 73.8, 71.8, 70.7, 70.0, 68.4, 67.3, 66.4, 55.2, 53.5, 48.1, 43.4, 42.1, 39.2, 39.1, 38.5, 36.3, 35.4, 35.1, 33.7,

33.3, 32.8, 32.2, 29.8, 29.7, 24.8, 24.6, 22.4, 21.2, 21.1, 21.0, 18.9, 17.5, 14.8, 14.2, 13.6, 9.9; HRMS calcd for $C_{54}H_{86}NaO_{13}~(M~+~Na)^+$ 965.5966, found 965.5897 (ESI).

(5S,7R,9S,12R,13S,14S)-15-((2S,3R,7R,E)-7-Acetoxy-2-((4S, 6R)-2,2-dimethyl-6-(pent-4-enyl)-1,3-dioxan-4-yl)-5-methy-Idec-4-en-3-yloxy)-13-(4-methoxybenzyloxy)-2,12,14-trimethyl-15-oxopentadec-1-ene-5,7,9-triyl Triacetate (34). To a solution of SI-4 (58 mg, 0.062 mmol) in CH_2Cl_2 (3.0 mL) were added DMAP (1 crystal), pyridine (0.2 mL, 2.46 mmol), and acetic anhydride (0.117 mL, 1.23 mmol). The reaction was stirred at rt until disappearance of starting material (~ 2 h). The reaction was diluted with EtOAc (3 mL) and quenched with saturated NH₄Cl (3 mL), and the aqueous layer was re-extracted with EtOAc (3 \times 5 mL). The organic layer was washed with brine (5 mL), dried (Na₂SO₄), filtered, and concentrated under reduced pressure. Flash chromatography (1.5:1 hexanes/EtOAc) provided 34 (60 mg, 98%) as a clear oil: $[\alpha]_D = +2.2$ (*c* = 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.18 (d, J = 8.6 Hz, 2H), 6.83 (d, J = 8.6 Hz, 2H), 5.80 (dddd, J = 16.9, 10.2, 6.7, and 6.7 Hz, 1H), 5.67 (dd, J = 9.9, 5.6 Hz, 1H), 5.16 (d, J = 9.7 Hz, 1H), 5.00 (ddd, J = 17.1, 3.4, 1.6 Hz, 1H), 5.00–4.92 (m, 3H), 4.92–4.84 (m, 2H), 4.73 (s, 1H), 4.66 (s, 1H), 4.50 (d, J = 10.9 Hz, 1H), 4.35 (d, J = 10.9 Hz, 1H), 3.79 (s, 3H), 3.75–3.68 (m, 1H), 3.62–3.55 (m, 2H), 2.68 (dt, *J* = 16.0, 6.9 Hz, 1H), 2.15-1.87 (m, 8H), 2.06 (s, 3H), 2.04 (s, 3H), 2.02 (s, 3H), 2.00 (s, 3H), 1.80–1.20 (m, 20H), 1.77 (s, 3H), 1.73 (s, 3H), 1.33 (s, 3H), 1.27 (s, 3H), 1.06 (d, J = 7.1 Hz, 3H), 0.93–0.84 (m, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 175.1, 170.6, 170.6, 170.6, 170.5, 158.9, 144.7, 138.7, 138.6, 131.3, 128.8, 122.5, 114.6, 113.5, 110.3, 100.2, 83.2, 73.8, 72.1, 71.2, 70.7, 70.0, 67.3, 66.4, 55.2, 53.5, 44.2, 42.1, 39.2, 39.1, 35.9, 35.4, 33.7, 33.3, 32.2, 31.9, 31.6, 29.9, 24.8, 24.8, 22.7, 21.3, 21.2, 21.1, 21.0, 18.4, 17.8, 14.7, 14.2, 14.2, 14.0, 13.2, 9.7; HRMS calcd for $C_{56}H_{88}NaO_{14} (M + Na)^+$ 1007.6072, found 1007.6210 (ESI).

(5S,7R,9S,12R,13S,14S)-15-((4R,8R,9S,10S,12R,E)-4-Acetoxy-10,12-dihydroxy-6,9-dimethylheptadeca-6,16-dien-8-yloxy)-13-(4-methoxybenzyloxy)-2,12,14-trimethyl-15-oxopentadec-1-ene-5,7,9-triyl Triacetate (SI-5). To a solution of tetraacetate 34 (60 mg, 0.061 mmol) in MeOH (6 mL) was added PPTS (2.5 mg, 0.03 mmol). The reaction was stirred until disappearance of starting material at rt (\sim 4 h). The reaction was diluted with EtOAc and quenched with saturated NaHCO₃, and the aqueous layer was re-extracted with EtOAc (3 \times 10 mL). The organic layer was washed with brine (10 mL), dried (Na₂SO₄), filtered, and concentrated under reduced pressure. Flash chromatography (1.5:1 hexanes/EtOAc) provided SI-5 (47 mg, 82%) as a clear oil: $[\alpha]_D = +8.97$ (c = 1.7, CHCl₃); ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3) \delta 7.21 (d, J = 8.3 \text{ Hz}, 2\text{H}), 6.85 (d, J = 8.4 \text{ Hz}, 2\text{H}), 5.82$ (dddd, J = 15.8, 12.1, 5.4, 5.4 Hz, 1H), 5.69 (dd, J = 9.7, 6.0 Hz, 1H), 5.18 (d, J = 9.8 Hz, 1H), 5.05-4.85 (m, 6H), 4.73 (s, 1H), 4.67 (s, 1H), 4.51 (d, J = 10.9 Hz, 1H), 4.34 (d, J = 11.0 Hz, 1H), 3.92–3.86 (m, 1H), 3.80 (s, 3H), 3.71 (m, 1H), 3.58 (dd, J = 8.4, 2.0 Hz, 1H), 2.73 (dddd, J = 14.2, 6.8, 6.8, 6.8 Hz, 1H), 2.50-1.19 (m, 28H), 2.07 (s, 3H), 2.03 (s, 3H), 2.02 (s, 3H), 1.82 (s, 3H), 1.72 (s, 3H), 1.09 (d, J = 7.0 Hz, 3H), 0.89 (t, J = 7.3 Hz, 3H), 0.86–0.81 (m, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 175.1, 170.7, 170.6, 170.6, 170.5, 158.9, 144.7, 138.7, 138.6, 131.1, 128.4, 123.4, 114.7, 113.7, 110.3, 83.5, 73.8, 73.5, 72.5, 70.7, 70.1, 68.8, 67.3, 60.4, 55.2, 44.3, 43.4, 43.0, 39.5, 39.1, 38.5, 37.1, 36.3, 34.9, 33.7, 33.3, 32.2, 31.6, 29.8, 25.1, 22.7, 21.3, 21.2, 21.1, 21.1, 18.4, 17.8, 14.2, 14.0, 13.6, 10.8; HRMS calcd for $C_{53}H_{84}NaO_{14} (M + Na)^+$ 967.5759, found 967.5789 (ESI).

(55,7*R*,95,12*R*,135,145)-15-((4*R*,8*R*,95,105,12*R*,*E*)-4-Acetoxy-10,12-dihydroxy-6,9-dimethylheptadeca-6,16-dien-8-yloxy)-13-hydroxy-2,12,14-trimethyl-15-oxopentadec-1ene-5,7,9-triyl Triacetate (35). Ester SI-5 (105 mg, 0.112 mmol) was taken up in CH₂Cl₂ (5.0 mL) followed by the addition of pH = 7 buffer solution (5.0 mL) and DDQ (51 mg, 0.224 mmol) at rt. Upon completion (~0.5 h, monitored by TLC), CH₂Cl₂ (13 mL) was added followed by saturated NaHCO₃ (1 mL). The layers were separated, and the aqueous layer was extracted with CH_2Cl_2 (3 × 10 mL). The combined organic layers were washed with brine (10 mL), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure. Flash chromatography (2:1 hexanes/EtOAc) afforded 35 (89 mg, 97%) as a clear oil: $[\alpha]_D = +4.4 (c = 0.50, CHCl_3); {}^{1}H NMR (400 MHz,$ $CDCl_3$) δ 5.79 (dddd, *J* = 17.0, 10.2, 6.7, 6.7 Hz, 1H), 5.19 (dd, *J* = 9.7, 9.5 Hz, 1H), 5.07-4.87 (m, 6H), 4.73 (s, 1H), 4.66 (s, 1H), 4.15-4.09 (m, 2H), 3.96–3.89 (m, 1H), 3.71 (dd, J = 9.9, 1.2 Hz, 1H), 2.54 (dddd, J = 14.1, 7.0, 7.0, 7.0 Hz, 1H), 2.17–1.19 (m, 28H), 2.07 (s, 3H), 2.03 (s, 3H), 2.02 (s, 3H), 1.82 (s, 3H), 1.72 (s, 3H), 1.08 (d, J = 7.0 Hz, 3H), 0.89 (t, J = 7.3 Hz, 3H), 0.86–0.81 (m, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 174.0, 172.4, 170.8, 170.8, 170.6, 144.6, 138.7, 138.0, 125.3, 114.6, 110.4, 72.6, 72.6, 72.4, 71.9, 70.7, 68.1, 67.1, 66.9, 45.3, 44.2, 42.7, 39.0, 37.6, 37.3, 36.9, 36.0, 33.7, 33.6, 33.3, 32.3, 31.5, 29.0, 25.3, 22.4, 21.4, 21.3, 21.2, 21.1, 18.5, 17.8, 13.9, 13.6, 12.5, 9.5; HRMS calcd for $C_{45}H_{76}NaO_{13} (M + Na)^+$ 847.5184, found 847.5183 (ESI).

Dolabelide C (1). To a refluxing solution of ester 23 (70 mg, 0.085 mmol) in degassed CH₂Cl₂ (175 mL) was added Grubbs cat-B catalyst (8.0 mg, 0.0085 mmol). The reaction was refluxed 2 h with the addition of more catalyst (4.0 mg, 0.00425 μ mol) after 2 h. A third portion of catalyst (4.0 mg, 0.00425) was added after an additiona 2 h; the reaction was refluxed for 6 h (monitored by TLC and LC-MS). The solution was cooled to rt and concentrated under reduced pressure. The resultant residue was purified via flash chromatography through two sequential columns (8:1 CH₂Cl₂/acetone) and (5:1 pentane/EtOAc) to afford 1 (14.0 mg, 21% yield) as an analytically pure sample and its C14-C15 Zconfigured diastereomer (10.0 mg, 15% yield) (vide infra): $[\alpha]_D = +2.9$ $(c = 0.63, \text{CHCl}_3)$; ¹H NMR (500 MHz, pyridine- d_5)^{36b} δ 6.10–5.90 (br m, 1H), 5.70 (t, J = 9.3 Hz, 1H), 5.67 (s, 1H), 5.40 (d, J = 9.5 Hz, 1H),5.38-5.31 (m, 2H), 5.30-5.23 (m, 2H), 5.16-5.10 (m, 1H), 4.88-4.82 (m, 1H), 4.37-4.32 (m, 1H), 4.03 (br d, J = 9.1 Hz, 1H), 2.93-2.85 (m, 1H), 2.52-2.48 (m, 1H), 2.32 (dd, J = 14.0, 7.9 Hz, 1H), 2.28 (dd, J = 13.9, 5.4 Hz, 1H), 2.21–2.15 (m, 1H), 2.11–2.00 (m, 6H), 2.08 (s, 3H), 2.06 (s, 3H), 2.05 (s, 3H), 2.05 (s, 3H), 1.98 (s, 3H), 1.96-1.92 (m, 2H), 1.90-1.80 (m, 4H), 1.79-1.58 (m, 7H), 1.59 (s, 3H), 1.53–1.49 (m, 3H), 1.32–1.28 (m, 2H), 1.19 (d, J = 7.3 Hz, 3H), 1.16 $(d, J = 7.0 \text{ Hz}, 3\text{H}), 0.90 (d, J = 6.6 \text{ Hz}, 3\text{H}), 0.84 (t, J = 7.3 \text{ Hz}, 3\text{H}); {}^{13}\text{C}$ NMR (126 MHz, pyridine-*d*₅) δ 173.9, 170.6, 170.5, 170.4, 170.3, 136.7, 132.6, 127.3, 127.2, 74.3, 73.5, 71.8, 69.9, 69.9, 68.0, 67.9, 67.3, 46.4, 44.5, 43.7, 38.8, 38.5, 38.0, 37.2, 36.3, 35.2, 34.1, 31.8, 31.6, 29.3, 28.0, 27.0, 21.1, 21.0, 20.9, 20.9, 18.8, 17.6, 15.2, 14.0, 13.8, 12.6, 11.0; ¹H NMR (500 MHz, $CDCl_3$) δ 5.35 (t, J = 9.1 Hz, 1H), 5.10–5.05 (m, 2H), 5.04–5.00 (m, 1H), 4.98-4.92 (m, 1H), 4.88-4.82 (m, 2H), 4.08 (s, 1H), 3.93 (s, 1H), 3.57 (s, 1H), 3.24 (s, 1H), 2.60-2.54 (m, 1H), 2.54-2.47 (m, 1H), 2.25 (dd, J = 13.8, 7.1 Hz, 1H), 2.21 (dd, J = 14.1, 5.7 Hz, 1H), 2.07 (s, 3H), 2.03 (s, 3H), 2.02 (s, 3H), 2.01 (s, 3H), 1.90–1.83 (m, 2H), 1.81 (s, 3H), 1.78-1.64 (m, 3H), 1.64-1.62 (m, 4H), 1.58 (s, 3H), 1.56 (m, 9H), 1.43–1.27 (m, 5H), 1.25 (s, 3H), 1.23–1.20 (m, 1H), 1.07 (d, J = 7.1 Hz, 3H), 0.89 (t, J = 7.3 Hz, 3H), 0.84 (m, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 173.8, 171.2, 171.0, 170.7, 170.4, 137.9, 133.1, 126.1, 125.0, 74.6, 73.1, 72.2, 69.8, 69.2, 68.7, 68.1, 67.7, 45.1, 44.3, 42.7, 39.1, 36.8, 36.1, 35.1, 34.6, 31.9, 31.7, 29.7, 28.3, 26.7, 25.0, 21.2, 21.2, 21.2, 21.1, 18.5, 17.7, 15.2, 13.9, 13.6, 12.6, 10.6; HRMS calcd for $C_{43}H_{72}NaO_{13}$ (M + Na)⁺ 819.4871, found 819.4858 (ESI).

Non-Natural C14–**C15** *Z*-Isomer of 1: $[\alpha]_D = +10.0 (c = 0.30, CHCl_3);$ ¹H NMR (500 MHz, pyridine- d_5)^{36c} δ 6.28 (br s, 1H), 6.20–6.05 (br m, 1H), 5.85 (t, *J* = 9.4 Hz, 1H), 5.51 (d, *J* = 8.8 Hz, 1H), 5.48–5.40 (m, 2H), 5.38–5.29 (m, 2H), 5.27–5.18 (m, 1H), 4.88–4.84 (m, 1H), 4.53–4.46 (m, 1H), 4.14 (br d, *J* = 7.4 Hz, 1H), 3.03–2.97 (m, 1H), 2.64–2.56 (m, 1H), 2.43 (dd, *J* = 7.5, 13.8 Hz, 1H), 2.37 (dd, *J* = 5.3, 13.1 Hz, 1H), 2.34–2.23 (m, 3H), 2.22–2.19 (m, 1H), 2.18–2.14 (m, 12H), 2.13–2.12 (m, 2 H), 2.08 (s, 3 H), 2.06–2.02 (m, 1 H), 2.02–2.00 (m, 1 H), 1.99–1.95 (m, 2 H), 1.95–1.91 (m, 1 H), 1.91–1.86 (m, 2 H), 1.84–1.78 (m, 2 H), 1.77 (s, 3 H), 1.75–1.66 (m, 3

H), 1.64–1.58 (m, 3 H); 1.49–1.31 (m, 4 H), 1.29 (d, J = 7.0 Hz, 6 H), 1.04 (d, J = 6.6 Hz, 3 H), 0.94 (t, J = 7.3 Hz, 3 H); ¹³C NMR (126 MHz, pyridine- d_5) δ 174.2, 170.9, 170.9, 170.7, 170.6, 137.2, 134.3, 127.3, 126.9, 75.2, 73.6, 72.2, 71.8, 70.1, 68.3, 68.2, 68.1, 46.6, 44.8, 44.3, 39.9, 39.2, 39.0, 37.6, 36.7, 34.4, 32.8, 32.0, 29.3, 28.4, 27.9, 27.5, 23.3, 21.4, 21.4, 21.3, 21.3, 19.1, 17.9, 14.3, 14.3, 13.0, 11.4; $^1{\rm H}$ NMR (500 MHz, CDCl₃) 36d δ 5.13-5.07 (m, 2H), 5.03-4.97 (m, 3H), 4.96-4.91 (m, 1H), 4.90-4.85 (m, 1H), 4.27 (s, 1H), 4.05–4.01 (m, 1H), 3.68 (d, J = 9.7 Hz, 1H), 2.54 (tt, J = 7.0 Hz, 1H), 2.31–2.20 (m, 4H), 2.13–2.11 (m, 1H), 2.11–2.08 (m, 1H), 2.06–2.04 (m, 2H), 2.04–2.03 (m, 6H), 2.02 (s, 3H), 2.01 (s, 3H), 2.00-1.97 (m, 1H), 1.97-1.95 (m, 1H), 1.82 (s, 3H), 1.81-1.76 (m, 2H), 1.76-1.74 (m, 1H), 1.74-1.71 (m, 1H), 1.66 (s, 3H), 1.61 (s, 1H), 1.58-1.56 (m, 1H), 1.48-1.46 (m, 2H), 1.45-1.43 (m, 2H), 1.42–1.39 (m, 2H), 1.38–1.25 (m, 6H), 1.01 (d, J=7.0 Hz, 3H), 0.89 (t, J= 7.3 Hz, 3H), 0.82 (d, J = 7.0 Hz, 3H), 0.79 (d, J = 6.7 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 173.6, 171.6, 171.3, 170.7, 170.4, 137.6, 133.9, 125.9, 125.8, 74.0, 72.8, 72.3, 71.3, 70.8, 68.0, 67.0, 66.6, 45.7, 44.1, 42.9, 38.0, 37.4, 37.0, 35.9, 34.9, 33.6, 32.2, 31.1, 28.2, 26.7, 25.9, 22.8, 21.4, 21.2, 21.2, 21.2, 21.2, 18.5, 17.8, 13.9, 13.4, 11.5, 9.36; HRMS calcd for $C_{43}H_{72}NaO_{13} (M + Na)^+ 819.4871$, found 819.4877 (ESI).

HPLC-MS Analysis of 1. HPLC data was collected using the following gradient over 35 min:

	A (%)	B (%)	
time	(99:1 H ₂ O/	(99:1 MeCN/	flow rate
(min)	MeCN)	$H_2O)$	(mL/min)
0.00	95.0	5.0	1.000
1.00	40.0	60.0	1.000
31.00	30.0	70.0	1.000

ASSOCIATED CONTENT

Supporting Information. Experimental details and spectroscopic data of new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

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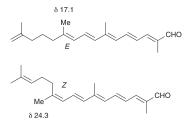
(31) Rigorous purification and degassing of the solvent was achieved through distillation over CaH_2 and a conventional freeze/thaw technique.

(32) The RCM was performed on 70 mg scale providing 14 mg of an analytically pure sample of the desired *E*-isomer and 10 mg of the *Z*-isomer.

(33) Optical rotation was measured over several trials resulting in variation of both the value and sign of analytically pure 1 (determined by LC–MS analysis, see the Supporting Information). This phenomenon is consistent with hydrogen-bonding systems, where inconsistency is frequently observed. Abraham, E.; Davies, S. G.; Roberts, P. M.; Russell, A. J.; Thomson, J. E. *Tetrahedron: Asymmetry* **2008**, *19*, 1027–1047 and references cited within.

(34) The major diastereomer was determined to be *E* due to resonances matching with listed resonances in reference.³ The *E* geometry is confirmed from comparison of ¹³C NMR, in which the C-14 methyl has a chemical shift of 15.7 ppm and the *Z*-isomer has a chemicals shift of 23.3 ppm, which is consistent with ref 3 and the paper cited therein (Carey, L.; Clough, J. M.; Pattenden, G. *J. Chem. Soc., Perkin Trans. 1* **1983**, 3005–3009) where it is stated that "The ¹³C NMR shifts

of vinyl methyl and vinyl methylene carbon atoms associated with isolated trisubstituted double bonds are critically dependent on the configuration of the double bond as a result of the well-known γ -effect." For example:



(35) Purification of each isomer was achieved using two to three consecutive runs on normal-phase flash chromatography (see the Supporting Information).

(36) (a) Spectrum integrated to 41 total H, where the unassigned H was presumed to be an O-H peak undergoing H-D exchange. (b) Spectrum integrated to 71 total H, where the unassigned H was presumed to be an O-H peak undergoing H-D exchange. Only the O-H peaks did not match exactly to the reported data. (c) Spectrum integrated to 71 total H, where the unassigned H was presumed to be an O-H peak undergoing H-D exchange. (d) Spectrum integrated to 69 total H, where the unassigned Hs were presumed to be O-H peaks undergoing H-D exchange.